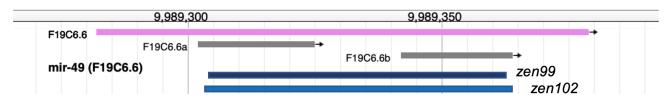


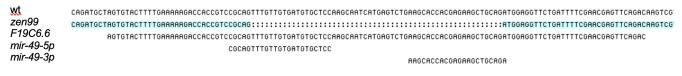
Two Deletion Alleles in the C. elegans mir-49 gene.

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Α



В



C

wt zen102	CAGATGCTAGTGTACTTTTGAAAAAGACCACCGTCCGCAGTTTGTTGTGATGTGCTCCAAGCAATCATGAGTCTGAAGCACCACGAGAAGCTGCAGATGGAGGTTCTGATTTTCGAACGAGTTCAGACGAGTTCAGACAAGTCG [CAGATGCTAGTGTACTTTTGAAAAAGACCACCGTCCGCA: ::::TGGAGGTTCTGATTTTCGAACGAGTTCAGACAAGTCG
F19C6.6	AGTGTACTTTTGAAAAAGACCACCGTCCGCAGTTTGTTGTGATGTGCTCCAAGCAATCATGAGTCTGAAGCACCACGAGAAGCTGCAGATGGAGGTTCTGATTTTCGAACGAGTTCAGAC
mir-49-5p	CGCAGTITGTTGTGTGTCCCC
mir-49-3p	AAGCACCACGAGAAGCTGCAGA

Figure 1: (A) A schematic of the *mir-49* locus and the location of the newly generated *mir-49*(*zen99*) and *mir-49*(*zen102*) deletion alleles. **(B)** *zen99* removes 56 base pairs from the *mir-49* precursor. **(C)** *zen102* removes 58 base pairs from the *mir-49* precursor.

Description

MicroRNAs (miRNAs) are small, non-coding RNAs that post-transcriptionally repress gene expression (Gebert and MacRae, 2018). While many miRNA genes and their families have been analyzed for function (Miska *et al.* 2007, Alvarez-Saavedra and Horvitz 2010), there are microRNA genes for which loss of function alleles have not yet been generated. There are no available alleles for the *C. elegans mir-49* gene.

Using CRISPR-Cas9 genome editing, we generated two deletion alleles, *zen99* and *zen102*, that disrupt the *C. elegans mir-49* gene (Fig 1A). *mir-49(zen99)* and *mir-49(zen102)* delete 56 base pairs and 58 base pairs from the *mir-49* locus, respectively (Fig 1B and Fig 1C). Each deletion nearly completely removes both strands generated by the *mir-49* locus, *mir-49-3p* and *mir-49-5p*. Both *mir-49* alleles are homozygous viable and appear to be superficially wild type. Careful phenotypic analysis will be important to characterize the effects of the two *mir-49* deletions.

Methods

Request a detailed protocol

To generate the *mir-49* deletion alleles, N2 animals were injected with the CRISPR-Cas9 components as an RNA-protein complex (Paix *et al.* 2015). The following components were used: Alt-R Cas9 (IDT, cat# 1081058) loaded with *mir-49* crRNAs (IDT, custom) (*mir-49* crRNA1 sequence: 5'-GAGCACACAAACTG-3', *mir-49* crRNA2 sequence: 5'-GCACCACGAGAAGCTGCAGA-3'), *dpy-10* targeting guide RNA (IDT, custom) (5'-GCUACCAUAGGCACCACGAG-3', Arribere *et al.* 2014) and tracer RNA (IDT, cat# 1072532) (AGCAUAGCAAGUUAAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUU).

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Briefly, to load the Alt-R Cas9, the following mixture was incubated at 37°C for 15 minutes: 0.5μL of Alt-R Cas9, 2.4μL of tracrRNA (0.4μg/μL), 0.8μL of *mir-49* crRNA1 (0.4μg/μL), 0.8μL of *mir-49* crRNA2 (0.4μg/μL), 1.3 μL of *dpy-10* crRNA (0.1μg/μL), 1μL IDT annealing buffer (provided with Alt-R Cas9), and 3.2μL of water. Following the incubation, the mixture was spun for 2 minutes at top speed (~10,000rpm). The progeny of the injected animals was first screened for the presence of dumpy worms to identify parents positive for Cas9 activity (Arribere *et al.* 2014). F1 offspring of the Cas9-positive parents were then genotyped for the presence of potential *mir-49* deletions using the following primers: mir-49.for1 (5'-AGGCACCACCTTACCATTCAT-3') and mir-49.rev1 (5'-GATGACTTACAGTCGCGTCTT-3'), which generate a wild type product of ~430 bps. Independent *mir-49* deletions were identified, homozygosed, and sequenced. The resultant strains, UY264 (*mir-49*(*zen99*)) and UY267 (*mir-49* (*zen102*)) were not outcrossed, but appear to be free of background *dpy-10* mutations. Sequencing was repeated in the next generation to ensure the stability of the generated alleles.

Reagents

UY264 mir-49(zen99) and UY267 mir-49 (zen102) are available upon request.

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