

Genetic Mapping of a new *Hippo* allele, *Hpo*^{N.1.2}, in *Drosophila melanogaster*

Jamie L Siders¹, Kayla L Bieser², Danielle R Hamill³, Erika C Acosta², Olivia K Alexander¹, Humza I Ali⁴, Micah J Anderson¹, Hayden R Arrasmith¹, Mustafa Azam⁴, Nikki J Beeman², Hassan Beydoun⁴, Lauren J Bishop¹, Morgan D Blair¹, Brianna Bletch⁴, Heather R Bline¹, Jennifer C Brown¹, Kelly M Burns¹, Karina C Calagua², Lexie Chafin³, William AH Christy³, Carlyn Ciamacco¹, Hannah Cizauskas⁴, Caitlyn M Colwell³, Abigail R Courtright³, Lucero Diaz Alavez², Rayne IS Ecret², Fatima Edriss⁴, Taylor G Ellerbrock¹, Madison M Ellis¹, Erica M Extine¹, Eric Feldman², Luke J Fickenworth¹, Caroline M Goeller¹, Alexis S Grogg¹, Yailine Hernandez², Abigail Hershner¹, Megan M Jaus¹, Leyre Jimenez Garcia¹, Katey E Franks¹, Ethan T Kazubski¹, Emily R Landis¹, Jon Langub², Tia N Lassek², Triet C Le¹, Julia M Lee¹, Daniel P Levine², Phoebe J Lightfoot³, Natasha Love², Ali Maalhigh-Fard⁴, Colin Maguire¹, Brynna E McGinnis², Bhargavi V Mehta¹, Veronica Melendrez², Zimri E Mena², Seth Mendell⁴, Petra Montiel-Garcia², Autumn S Murry^{4,5}, Riley A Newland¹, Ryan M Nobles², Neha Patel², Yashodhara Patil⁴, Cassidy L Pfister¹, Victoria Ramage¹, Mya R Ray¹, Joseph Rodrigues², Victoria C Rodriguez², Yara Romero², Alexandra M Scott¹, Nicholas Shaba⁴, Samantha Sieg⁴, Kayla Silva¹, Sahiba Singh⁴, Aleksandria J Spargo¹, Savanna J Spitnale¹, Nicole Sweeden², Logan Tague⁴, Breanna M Tavernini², Kathleen Tran¹, Liselle Tungol⁴, Kylie A Vestal¹, Amber Wetherbee¹, Kayla M Wright¹, Anthony T Yeager¹, Rehab Zahid⁴ and Jacob D Kagey^{4§}

¹School of Science, Technology, and Mathematics, Ohio Northern University, Ada, OH USA

²Department of Physical and Life Sciences, Nevada State College, Henderson, NV USA

³Department of Zoology, Ohio Wesleyan University, Delaware, OH USA

⁴Biology Department, University of Detroit Mercy, Detroit, MI USA

⁵ReBUILDetroit, University of Detroit Mercy, Detroit, MI USA

[§]To whom correspondence should be addressed: kageyja@udmercy.edu

Abstract

Genetic screens provide a mechanism to identify genes involved with different cellular and organismal processes. Using a Flp/FRT screen in the *Drosophila* eye we identified mutations that result in alterations and de-regulation of cell growth and division. From this screen a group of undergraduate researchers part of the Fly-CURE consortium mapped and characterized a new allele of the gene *Hippo*, *Hpo*^{N.1.2}.

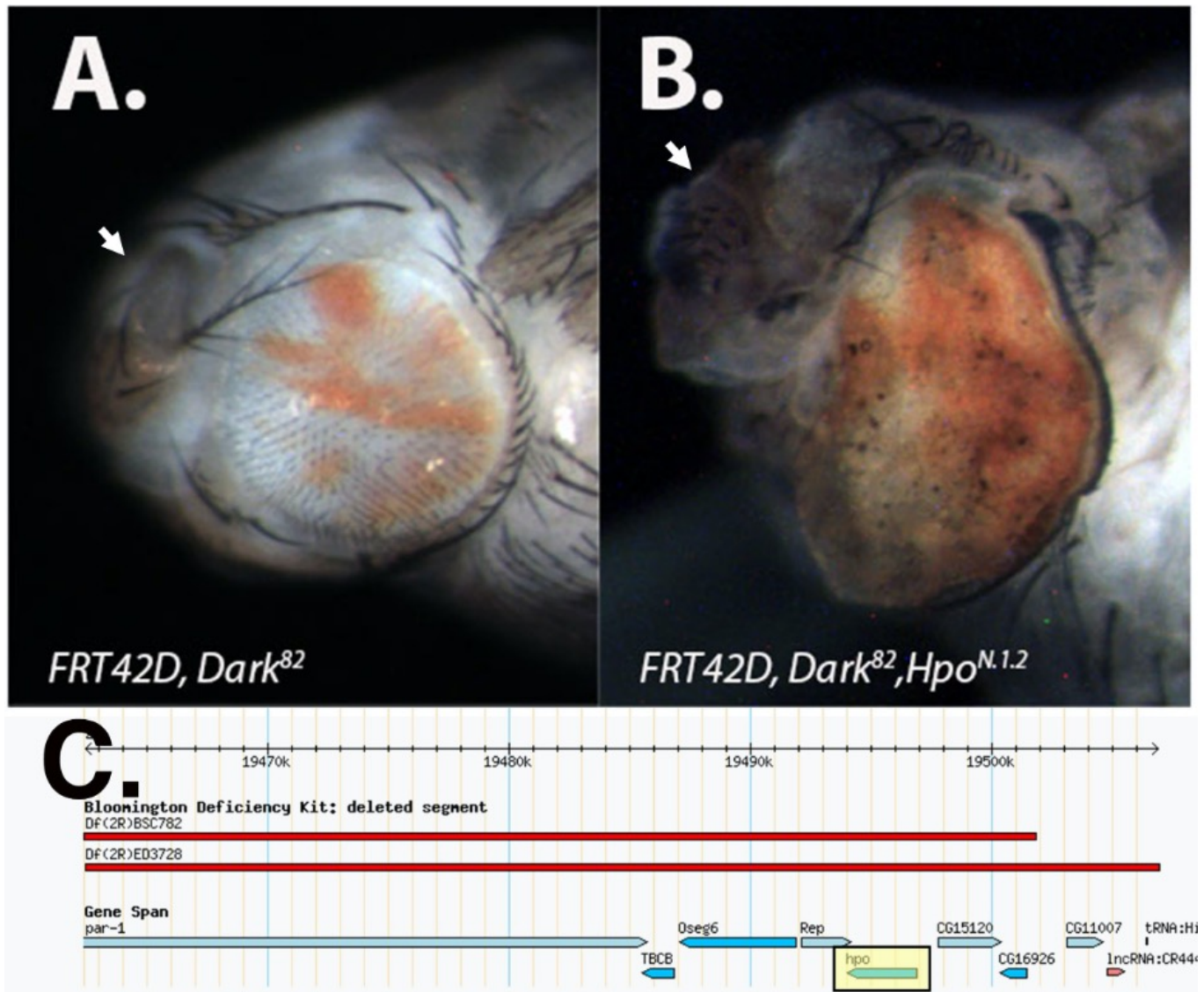


Figure 1. The *Dark⁸², Hpo^{N.1.2}* mosaic eye results in dramatic tissue overgrowth and pupal lethality. : A.) Lateral view of *FRT42D, Dark⁸²* mosaic control eye and B.) *FRT42D, Dark⁸², Hpo^{N.1.2}* mosaic mutant pupal eyes at the same magnification (40x). In both genotypes, mutant tissue displays red pigmentation (w^{+mC}). Arrows on A and B point to antennae, eyes are oriented with anterior to the left and dorsal to the top. C.) Genomic region of chromosome 2R in which mutant *N.1.2* failed to complement (2R:19,462,450..19,506,861). The *N.1.2* mutation mapped to the *hpo* gene locus found in this region (yellow highlight). Image adapted from flybase.org (Gramates 2017).

Description

In order to identify conditional regulators of cell growth and tumorigenesis, an EMS-based genetic screen was conducted in *Drosophila melanogaster* utilizing the Flp/FRT system in an apoptotic null background (Akdemir *et al.*, 2006). A fly line possessing the *FRT42D, Dark⁸²* chromosome and containing a mini-white cassette (w^{+mC}) was used for EMS mutagenesis. Subsequent matings to *FRT42D; Ey-Flp* flies facilitated phenotypic screening in eye tissue. Phenotypes observed in the screen included alterations in mosaicism (red > white pigmentation pattern), eye and/or antennal overgrowth, defects in patterning, and pupal lethality (Kagey *et al.* 2012). One of the pupal lethal mutants identified in this screen, *N.1.2*, is discussed here. Genetic crosses between *FRT42D, Dark⁸², N.1.2* X *FRT42D; Ey-Flp* resulted in near complete pupal lethality (~ 90%) due to dramatic tissue overgrowth of the eye, antennae, and inter-ocular space (Figure 1B compared to 1A). Due to the pupal lethality associated with the eye tissue overgrowth, flies were dissected from late pupal stages in order to visualize the phenotype. Imaging following dissection shows that the mutant eyes are comprised of mostly mutant (pigmented) tissue that protrudes

from the eye cavity creating tissue folding (Figure 1B). Control *FRT42D*, *Dark⁸²* flies exhibited a balance in the red:white ratio and no eye overgrowth (Figure 1A). For direct comparison to *N.1.2*, the *FRT42D*, *Dark⁸²* mosaic eye was also imaged at the late pupal stage. Eyes were imaged under 70% ethanol on an AM Scope digital camera at 40x using a LED light ring.

In order to map the genetic location of the *N.1.2* mutation, we conducted complementation tests with deficiency strains, and looked for lethality, consistent with the phenotype of homozygous mutant flies of the *N.1.2* stock. Deficiency crosses were conducted by mating *FRT42D*, *Dark⁸²*, *N.1.2/CyO* virgin females to males from each of 86 deficiency stocks with deletions distal to the *FRT42D* site on the right arm of chromosome two. All stocks used for mapping were part of the Bloomington Deficiency 2R Kit (Cook et al., 2012). The genetic mapping was conducted by undergraduate research students from Nevada State College, Ohio Northern University, Ohio Wesleyan University, and the University of Detroit Mercy as part of the Fly-CURE consortium (Vrailas-Mortimer et al. 2021; Bieser et al. 2019; Stamm et al. 2019). The *N.1.2* mutant failed to complement two of these deficiencies, *Df(2R)BSC782* (2R: 19,451,027..19,501,804) and *Df(2R)ED3728* (2R:19,462,450..19,726,747). These two deficiencies overlapped from cytological bands 56D10-56D14 (2R:19,462,450..19,506,861) and included the hippo (*hpo*) gene locus and nine other protein coding genes (2R:19,492,996..19,496,856) (Figure 1C). Subsequent crosses to individual lethal alleles of *hpo*: *hpo^{MGH1}* and *hpo^{KS240}*, failed to complement *N.1.2*, confirming that the *N.1.2* mutant is a newly isolated *hpo* allele, *hpo^{N.1.2}* (Harvey et al., 2003; Udan et al., 2003).

The *hpo* gene functions as a negative regulator of cell growth and is a part of a highly conserved signaling pathway first characterized in *Drosophila* (reviewed in Harvey and Hariharan 2012). Overall, research has shown that the hippo pathway is critical for regulating organ size through regulation of apoptosis, cell survival, cell polarity, and cell proliferation (Harvey et al., 2003; Jia et al. 2003; Pantalacci et al., 2003; Udan et al., 2003; reviewed in Yu et al., 2015). Consistent with the eye phenotypes observed in the *N.1.2* mutation, mutations in *hpo* are known to result in striking overgrowth phenotypes in a variety of tissue types from flies to humans (Pan 2010, Pluoffe et al., 2015; Yu et al., 2015). Identification of this novel *hpo^{N.1.2}* allele will support further research into the molecular mechanisms by which multicellularity is regulated and restricted by this critical signaling pathway.

Reagents

FRT42D, *Dark⁸²/CyO* (Akdemir et al., 2006)

FRT42D, *Dark⁸²*, *hpo^{N.1.2}/CyO* (this manuscript)

FRT42D; *Ey-Flp* (BDSC 8211)

Bloomington *Drosophila* Stock Center 2R Deficiency Kit (Cook et al., 2012)

w¹¹¹⁸; *Df(2R)BSC782/SM6a* (BDSC 27354)

w¹¹¹⁸; *Df(2R)ED3728/SM6a* (BDSC 9067)

yw; *FRT42D*, *hpo^{KS240}/CyO* (BDSC 25085)

hpo^{MGH1} (Harvey et al. 2003)

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Author Contributions: Jamie L Siders: Data curation, Formal analysis, Investigation, Project administration, Supervision, Validation, Writing - original draft, Writing - review and editing. Kayla L Bieser: Data curation, Formal analysis, Funding acquisition, Investigation, Project administration, Resources, Software, Supervision, Validation, Writing - review and editing. Danielle R Hamill: Data curation, Formal analysis, Investigation, Project administration, Resources, Supervision, Validation, Writing - review and editing. Erika C Acosta: Data curation, Formal analysis, Investigation, Writing - review and editing. Olivia K Alexander: Data curation, Formal analysis, Investigation, Writing - review and editing. Humza I Ali: Data curation, Formal analysis, Investigation, Writing - review and editing. Micah J Anderson: Data curation, Formal analysis, Investigation, Writing - review and editing. Hayden R Arrasmith: Data curation, Formal analysis, Investigation, Writing - review and editing. Mustafa Azam: Data curation, Formal analysis, Investigation, Writing - review and editing. Nikki J Beeman: Data curation, Formal analysis, Investigation, Writing - review and editing. Hassan Beydoun: Data curation, Formal analysis, Investigation, Writing - review and editing. Lauren J Bishop: Data curation, Formal analysis, Investigation, Writing - review and editing. Morgan D Blair: Data curation, Formal analysis, Investigation, Writing - review and editing. Brianna Bletch: Data curation, Formal analysis, Investigation, Writing - review and editing. Heather R Bline: Data curation, Formal analysis, Investigation, Writing - review and editing. Jennifer C Brown: Data curation, Formal analysis, Investigation, Writing - review and editing. Kelly M Burns: Data curation, Formal analysis, Investigation, Writing - review and editing. Karina C Calagua: Data curation, Formal analysis, Investigation, Writing - review and editing. Lexie Chafin: Data curation, Formal analysis, Investigation, Writing - review and editing. William AH Christy: Data curation, Formal analysis, Investigation, Writing - review and editing. Carlyn Ciamacco: Data curation, Formal analysis, Investigation, Writing - review and editing. Hannah Cizauskas: Data curation, Formal analysis, Investigation, Writing - review and editing. Caitlyn M Colwell: Data curation, Formal analysis, Investigation, Writing - review and editing. Abigail R Courtright: Data curation, Formal analysis, Investigation, Writing - review and editing. Lucero Diaz Alavez: Data curation, Formal analysis, Investigation, Writing - review and editing. Rayne IS Ecret: Data curation, Formal analysis, Investigation, Writing - review and editing. Fatima Edriss: Data curation, Formal analysis, Investigation, Writing - review and editing. Taylor G Ellerbrock: Data curation, Formal analysis, Investigation,

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Ryan M Nobles: Data curation, Formal analysis, Investigation, Writing - review and editing. Neha Patel: Data curation, Formal analysis, Investigation, Writing - review and editing. Yashodhara Patil: Data curation, Formal analysis, Investigation, Writing - review and editing. Cassidy L Pfister: Data curation, Formal analysis, Investigation, Writing - review and editing. Victoria Ramage: Data curation, Formal analysis, Investigation, Writing - review and editing. Mya R Ray: Data curation, Formal analysis, Investigation, Writing - review and editing. Joseph Rodrigues: Data curation, Formal analysis, Investigation, Writing - review and editing. Victoria C Rodriguez: Data curation, Formal analysis, Investigation, Writing - review and editing. Yara Romero: Data curation, Formal analysis, Investigation, Writing - review and editing. Alexandra M Scott: Data curation, Formal analysis, Investigation, Writing - review and editing. Nicholas Shaba: Data curation, Formal analysis, Investigation, Writing - review and editing. Samantha Sieg: Data curation, Formal analysis, Investigation, Writing - review and editing. Kayla Silva: Data curation, Formal analysis, Investigation, Writing - review and editing. Sahiba Singh: Data curation, Formal analysis, Investigation, Writing - review and editing. Aleksandria J Spargo: Data curation, Formal analysis, Investigation, Writing - review and editing. Savanna J Spitale: Data curation, Formal analysis, Investigation, Writing - review and editing. Nicole Sweeden: Data curation, Formal analysis, Investigation, Writing - review and editing. Logan Tague: Data curation, Formal analysis, Investigation, Writing - review and editing. Breanna M Tavernini: Data curation, Formal analysis, Investigation, Writing - review and editing. Kathleen Tran: Data curation, Formal analysis, Investigation, Writing - review and editing. 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