Genome Sequence of *Microbacterium foliorum* Phage KingKamren

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Abstract

We report the discovery and genome sequence of a cluster EK bacteriophage, KingKamren, isolated from a soil sample collected in Plattsburgh, New York using the bacteria *Microbacterium foliorum*, B-24224. Its 54,721 bp genome contains 51 putative genes, 17 of which have predicted functions.

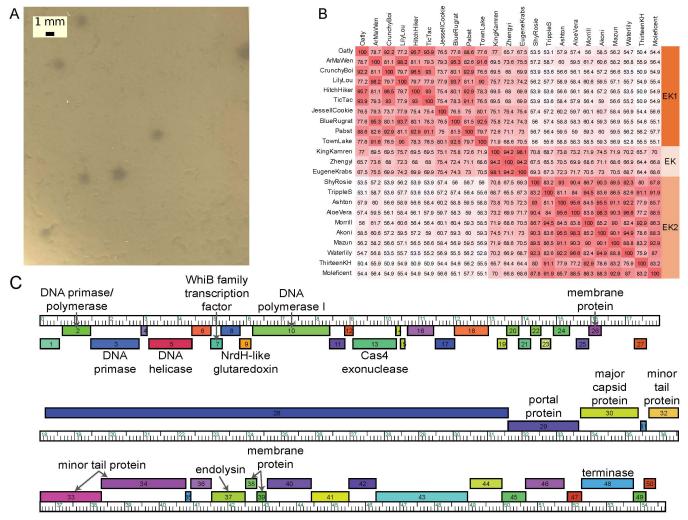


Figure 1. KingKamren plaques, Gene Content Similarity Comparison, and genome:

(A) Plaques of phage KingKamren in top agar with *M. foliorum*. (B) Gene Content Similarity map, containing GSC values from the GCS tool (Hirokawa et al., 1998) using random selection of 10 phages from the EK1 and EK2 subclusters and all current EK cluster phages. (C) Genome map for KingKamren, with putative genes presented as colored boxes along a genome ruler, in kilobases. Boxes above and below the ruler represent genes that are transcribed rightwards and leftwards, respectively.

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Description

Bacteriophages are some of the most genetically diverse entities known (Pope et al., 2015). Understanding this diversity has implications for both ecological and health applications (Milhaven et al., 2023; Strathdee et al., 2023). Here we present the genome of a new bacteriophage, KingKamren, which was collected in 2023 from Plattsburgh NY (44.69179 N, 73.46351 W) using the bacterial host *Microbacterium foliorum*, B-24224.

Following standard procedures (Zorawik et al., 2024), approximately 15 cm³ of soil was suspended in 35 mL PYCa medium and shaken at 37°C at 250 rpm for 2 hours followed by centrifugation at 2,000 g and vacuum filtration (0.22 μ m filter) of the supernatant. This filtrate was inoculated with *M. foliorum* and incubated at 30°C for 5 days at 250 rpm. An aliquot was spun at 14,000 g, filtered, and plated in PYCa top agar containing *M. foliorum*. After 48 hours, KingKamren formed small, turbid plaques with an average size of 1.17 mm (\mp 0.10 SE) in diameter, as determined through measurements using ImageJ (Schnieder et al., 2012) (Fig. 1A) and was purified through two rounds of plating.

DNA was isolated from a lysate (Wizard DNA Clean-up Kit, Promega), prepped for sequencing (NEBNext Ultrall FS Kit), sequenced (Illumina sequencing, v3 reagents) and assembled as described by Russell (Russell, 2018). Sequencing resulted in 2,929,352 single-end 150-bp reads with 4,991-fold coverage. The genome was assembled using Newbler v2.9 (Margulies et al., 2005) and checked for completeness and genome termini using Consed v29.0 (Gordon et al., 1998). Default settings were used unless otherwise noted. This resulted in a genome 54,721 bp in length with 203-bp direct terminal repeat ends and a GC content of 57.5%.

Using standard procedures (Pope et al., 2017), the software DNA Master v5.23.6 (http://cobamide2.bio.pitt.edu), PECAAN (https://discover.kbrinsgd.org), Genemark v2.5p (Lukashin and Borodovsky, 1998), and Glimmer v3.02 (Delcher et al., 1999) were used to predict 53 protein-encoding genes. Start sites were determined using Starterator v485 (https://seaphages.org/software/#Starterator) and Blastp v2.13.0 (Altschul et al., 1990) alignments against the Actinobacteriophage protein (Russell and Hatful, 2017) and NCBI non-redundant protein sequences databases (https://blast.ncbi.nlm.nih.gov). No strong evidence for tRNAs was found using Aragorn v1.2.41 (Laslett and Canback, 2004) and tRNAscan-SE v2.0 (Lowe and Eddy, 1997). A total of 14 genes were assigned putative functions using BLASTp v2.13.0 (Altschul et al., 1990), Phamerator (Cresawn et al., 2011), and HHpred (searching against PDB_mmCIF70, SCOPe70, Pfam-A, and NCBI_Conserved_Domains databases) (Söding et al., 2005). deepTMHMM v1.0.24 (Krogh et al., 2001) and SOSUI (Hirokawa et al. 1998) detected an additional 3 genes as membrane proteins. All software used default settings. The annotation is presented in Fig. 1B

KingKamren was assigned to the EK cluster using the GCS tool (Hirokawa et al., 1998), based on having a gene content similarity (GCS) of at least 35% to other EK bacteriophages in the Actinobacteriophage database. The EK cluster currently contains 56 members. The majority of EK phages are placed into one of two subclusters, EK1 or EK2, but KingKamren is one of three phages (to date) that are not sub-classified, as its GCS is similar to both EK1 and EK2 phages (Figure 1b). This small subset of EK phages share 4 genes (KingKamren's genes 12, 26, 35, and 47 – Fig. 1C) of unknown function that are unique to this group and do not share significant sequence similarity to any other actinobacteriophage in the database (phagesdb.org). KingKamren also has a 13,452 bp gene (gene 28, Fig. 1B) which constitutes 24.6% of its entire genome and encodes a 4,483 amino acid protein of unknown function. This feature is found across all members in the EK phage cluster and represents one of the largest genes in actinobacteriophages (Jacobs-Sera et al., 2020).

Nucleotide sequence accession numbers

KingKamren is available at GenBank with Accession No. XPP978791 and Sequence Read Archive (SRA) No. SRX25029057.

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References

Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/S0022-2836(05)80360-2 DOI: <u>https://doi.org/10.1016/S0022-2836(05)80360-2</u>

Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. BMC Bioinformatics 12:395–395. https://doi.org/10.1186/1471-2105-12-395



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Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. Nucleic Acids Res 27:4636–4641. https://doi.org/10.1093/nar/27.23.4636 DOI: <u>10.1093/nar/27.23.4636</u>

Hirokawa T, Boon-Chieng S, Mitaku S. 1998. SOSUI: classification and secondary structure prediction system for membrane proteins. Bioinformatics 14:378–379. https://doi.org/10.1093/bioinformatics/14.4.378

Gordon D, Abajian C, Green P. 1998. *Consed:* A Graphical Tool for Sequence Finishing. Genome Research 8: 195-202. DOI: <u>10.1101/gr.8.3.195</u>

Jacobs-Sera D, Abad LA, Alvey RM, Anders KR, Aull HG, Bhalla SS, . Blumer LS, Bollivar DW, Bonilla JA, Butela KA, Coomans RJ, Cresawn SG, D'Elia T, Diaz A, Divens AM, Edgington NP, Frederick GD, Gainey MD, Garlena RA, Grant KW, Gurney SMR, Hendrickson HL, Hughes LE, Kenna MA, Klyczek KK, Kotturi HK, Mavrich TN, McKinney AL, . Merkhofer EC, Parker JM, . Molloy SD, Mont DLi, Pape-Zambito DA, Pollenz RS, Pope WH, Reyna NS, Rinehart CA, Russell DA, . Shaffer CD, Sivanathan V, Stoner TH, Stukey JS, Sunnen CN, Tolsma SS, Tsourkas PK, Wallen JR, Ware VC, Warner MH, Washington JM, Westover KM, Whitefleet-Smith JL, Wiersma-Koch HI, Williams DC, Zack KM, Hatfull GF. 2020. Genomic diversity of bacteriophages infecting *Microbacterium* spp. PLOS One. https://doi.org/10.1371/journal.pone.0234636

Krogh A, Larsson B, von Heijne G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. J Mol Biol 305:567–580. https://doi.org/10.1006/jmbi.2000.4315

Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152

Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25:955–964. https://doi.org/10.1093/nar/25.5.955

Lukashin AV, Borodovsky M. 1998. GeneMark.hmm: new solutions for gene finding. Nucleic Acids Res 26:1107–1115. https://doi.org/10.1093/nar/26.4.1107 DOI: <u>10.1093/nar/26.4.1107</u>

Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, et al., Rothberg JM. 2005. Genome sequencing in microfabricated high-density picolitre reactors. Margulies2005. DOI: <u>10.1038/nature03959</u>

Milhaven M, Versoza CJ, Garg A, Cai L, Cherian S, Johnson K, et al., Pfeifer SP. 2023. Microbacterium Cluster EA Bacteriophages: Phylogenomic Relationships and Host Range Predictions. Microorganisms. 11 DOI: 10.3390/microorganisms11010170

Pope WH, Bowman CA, Russell DA, Jacobs Sera D, Asai DJ, Cresawn SG, et al., Mycobacterial Genetics Course. 2015. Whole genome comparison of a large collection of mycobacteriophages reveals a continuum of phage genetic diversity. DOI: <u>10.7554/eLife.06416</u>

Pope WH, Jacobs-Sera D, Russell DA, Cresawn SG, Hatfull GF. 2017. SEA-PHAGES Bioinformatics Guide. Howard Hughes Medical Institute, Chevy Chase, MD.

Russell DA. 2018. Sequencing, Assembling, and Finishing Complete Bacteriophage Genomes. Methods Mol Biol. 1681:109-125. doi: 10.1007/978-1-4939-7343-9_9. PubMed ID: <u>29134591</u>

Russell DA, Hatfull GF. 2017. PhagesDB: the actinobacteriophage database. Bioinformatics 33:784–786. https://doi.org/10.1093/bioinformatics/btw711

Schneider CA, Rasband WS, Eliceiri KW. 2012. NIH Image to ImageJ: 25 years of image analysis. Nature Methods. 9: 671-675. DOI: <u>doi:10.1038/nmeth.2089</u>

Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. Nucleic Acids Res 33:W244–W248. https://doi.org/10.1093/nar/gki408

Strathdee SA, Hatfull GF, Mutalik VK, Schooley RT. 2023. Phage therapy: From biological mechanisms to future directions. Cell 186(1):17-31. doi: 10.1016/j.cell.2022.11.017. PubMed ID: <u>36608652</u>

Zorawik M, Jacobs Sera D, Freise AC, Reddi K. 2024. Isolation of Bacteriophages on Actinobacteria Hosts. Methods Mol Biol. 2793:273-298. doi: 10.1007/978-1-0716-3798-2_17. PubMed ID: <u>38526736</u>

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