

Precision deletion of the entire coding sequence of the *mod-5* locus causes increase in pharyngeal pumping frequency

Trisha Brock¹, Stelian Pop¹, Chandler Bradford¹, Jenn Lawson¹, Lauren Resch¹, Christopher Hopkins¹

¹Knudra Transgenics, 5201 S Green St., Salt Lake City, UT 84123, USA.

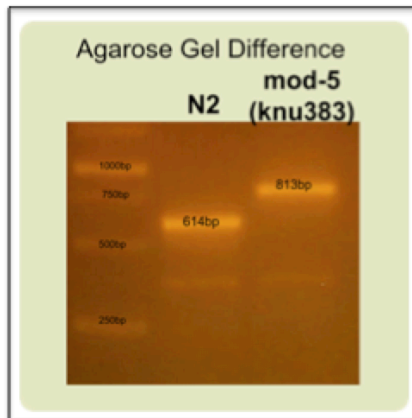


Figure 1A

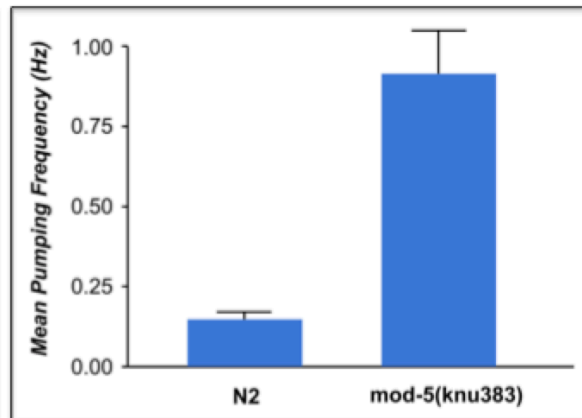


Figure 1B

Description

The *C. elegans* *mod-5* gene encodes a serotonin transporter. The putative null allele contains a 1,688bp deletion leading to a premature stop codon in the resulting sequence (Ranganathan, 2001); however, unspliced the gene is 12,557bp in length from the start to the stop codon. We used CRISPR/Cas9 technology to create a 12,775bp deletion that eliminates all of the *mod-5* coding sequence from the genome (Figure 1A).

The *mod-5(knu383)* animals were measured for pharynx pumping frequency using the NemaMetrix ScreenChip (<https://doi.org/10.17912/W2CC7Z>; protocol described here <http://www.knudra.com/p003>). Compared with N2, *mod-5(knu383)* worms showed a statistically significant increase in pumping frequency ($p < 0.0001$) (Figure 1B). For N2, 18 worms were tested in M9 with no stimulation, and 13 worms were tested for *mod-5(knu383)*.

Construction Details

The *mod-5(knu383)* allele repair oligonucleotide contained a 3-frame stop of TAAATAAATAAA surrounded by filler sequence of CCTCCCGTTCGCCTGGGACATC and GATGTCCAGGCCGAACGGGAGG. The homology arms were designed to give perfect homology for each of the sgRNA cut sites with 35 nucleotides of perfect homology on the 5' side and 34 nucleotides on the 3' side. These homology arms are 12,775bp apart from each other in the genomic sequence. The deletion was created using the CRISPR/Cas9 technology (Paix, 2014; Kim, 2014). The guide sequences were caaaagaaaagagcagccga and caaaagaaaagagcagccga provided in the injection mix as synthetic RNA. The deletion was detected by a three-primer PCR approach where an 813bp band would amplify in the deletion and a 614bp band would amplify in N2 wild-type, Figure 1A.

References

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Reagents

Strain: N2, COP1365 - *mod-5* (*knu383* [12.8 kb entire coding-sequence deletion]), NemaMetrix ScreenChip (Nemametrix)

Funding: Knudra Transgenics

Reviewed by: Peer reviewed

Received 05/22/2017, **Accepted** 05/31/2017. **Available** starting [WormBase](#) release WS261, **Published Online** 06/06/2017.

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Citation: Brock, T; Pop, S; Bradford, C; Lawson, J; Resch, L; Hopkins, C (2017): Precision deletion of the entire coding sequence of the *mod-5* locus causes increase in pharyngeal pumping frequency. *Micropublication:biology*. Dataset. <https://doi.org/10.17912/W2NP4D>.

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